## Magnetic properties of oxyhemoglobin

(hemoglobin/iron electronic state)

M. CERDONIO\*†‡, A. CONGIU-CASTELLANO§, F. MOGNO\*, B. PISPISA‡¶, G. L. ROMANI\*, AND S. VITALE\*

\* Department of Physics and \* Department of Physical Chemistry, University of Rome, Rome, Italy; and § Department of Physics, University of Cagliari, Cagliari, Italy

Communicated by Harry B. Gray, October 28, 1976

ABSTRACT When the magnetic susceptibility of frozen aqueous solutions of human oxyhemoglobin was measured in the range between 25 and 250 K, it showed a temperature-dependent behavior typical of a thermal equilibrium between a ground singlet state and an excited triplet state for two electrons per heme, the energy separation being  $|2J| = 146 \text{ cm}^{-1}$ . By contrast, within the same temperature range, carboxyhemoglobin was found to be diamagnetic, as already reported.

Since the first magnetic studies by Pauling and Coryell (1), the electronic state of iron and oxygen in oxyhemoglobin ( $HbO_2$ ) has been the subject of continuous debate. Five different models for  $O_2$  binding in  $HbO_2$  have been proposed on the basis of various theoretical interpretations of the available experimental evidence (2–6).

The presence of unpaired electrons in the iron-oxygen bond has been inferred from x-ray fluorescence spectra (7), from spectroscopy data and analogy with cobalt complexes (8, 9), and from Mössbauer (10) and Raman (11) spectra.

Despite this, in the majority of papers in this field, it is assumed that HbO<sub>2</sub> and carboxyhemoglobin (HbCO) are diamagnetic, which has precluded definite assignment of the electronic state of the site and has strongly biased the discussion

The present work is intended to remove this bias and to produce new experimental evidence about the magnetic state of HbO<sub>2</sub>, which is at variance with the commonly accepted behavior as cited above. Our experimental method, based on our oscillating sample (12) superconducting magnetometer (13), differs from previous methods mainly in the fact that we scan a wide temperature range, between 25 and 250 K, rather than take measurements only at a single temperature. This procedure allows us to resolve small paramagnetic contributions on top of the dominating background of the frozen protein solution without resorting to less direct subtraction procedures.

## MATERIALS AND METHODS

Native and stripped human  $\dot{H}bO_2$  was prepared according to the method of Benesch et al. (14). Sample concentrations were in the range of 10 mM in heme. Measurements always were made on fresh samples, by the following procedure. Within 24 hr after blood was drawn from one of us (F.M.), part of the sample at proper concentration and pH was saturated with air or pure  $O_2$  and brought to liquid helium temperature in the susceptometer, and its temperature-dependent susceptibility was measured. The rest of the sample was stored at 4 °C in equilibrium with air. The next day it was equilibrated with pure CO and then inserted in the susceptometer for measurements.

Optical spectra of the samples were recorded by a Beckman DK-2A spectrophotometer before and after the susceptibility measurement. The concentration of methemoglobin (MetHb) in the samples was estimated to be less than 3% in all cases. To avoid any possible interferences from MetHb, we added KCN to stripped HbO<sub>2</sub> to stabilize the MetHb in the low-spin form (15) at any temperature. Measurements were performed on several different preparations of Hb. The temperature-dependent susceptibility was measured at least twice on each sample.

## **RESULTS AND DISCUSSION**

To emphasize the small differences between HbO<sub>2</sub> and HbCO samples, all the data were normalized to give the same extrapolation for infinite temperature, which was set as the value extracted from the data of Havemann (16) for HbCO. Typical results for stripped HbO<sub>2</sub> and HbCO equilibrated with air, O<sub>2</sub> at 1 atm, and CO at 1 atm, are presented in Fig. 1.

There is a striking difference in behavior. The stripped HbCO showed a weak Curie law paramagnetic contribution which is quantitatively accounted for by the presence of 0.85% MetHb in the low-spin state.

Native HbCO gave similar results, with an even smaller paramagnetic contribution. We conclude that both native and stripped HbCO are diamagnetic. We also conclude that (i) no sample density effects interfered with the volume susceptibility measurements within our resolution and (ii) MetHb, if any, must stay in the same low-spin state in the whole temperature range scanned and strictly obey a simple Curie law.

The temperature dependence of the magnetic susceptibility of the frozen solutions of stripped HbO<sub>2</sub> was strikingly different from that of HbCO.

To fit the data we needed a composite temperature-dependent behavior that allows for the presence of a Curie law paramagnetic component plus a paramagnetic contribution nonlinear in inverse temperature, representing a thermal equilibrium between a ground singlet state and an excited triplet state. The Curie law paramagnetic contribution is easily assigned to the paramagnetism of the oxygen in solution and to an amount of low-spin MetHb smaller than 3%. To account for the nonlinear paramagnetic component we have assumed that two unpaired electrons are present in the iron-oxygen bond and their spins pair off in a singlet state at low temperature (<40 K) and thermally populate a triplet state at higher temperature. To describe this behavior we have used the appropriate theoretical relationships for two spins with  $S_1 = S_2 = \frac{1}{2}$  and singlet-triplet separation of -2J, as given in textbooks (17). The full curves in Fig. 1 represent a computer-performed least-square best fit to the data, with J and the Curie constant of the paramagnetic component left as free parameters. For the singlettriplet half separation of one spin pair per heme we get J = -72 $cm^{-1}$ 

<sup>†</sup> Present address: Faculty of Science, Free University of Trento, Povo, Trento, Italy.

<sup>&</sup>lt;sup>‡</sup> To whom correspondence should be addressed.

Present address: Laboratorio Elettronica Stato Solido, via Cineto Romano 42, Rome, Italy.

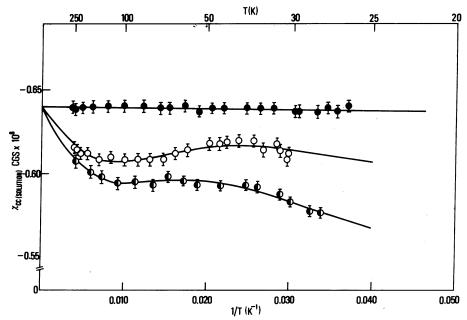


FIG. 1. Magnetic susceptibility of ligated stripped human hemoglobin plotted against reciprocal of inverse temperature, shown as mean. O, HbO<sub>2</sub> equilibrated with air, pH 7.2; heme concentration 9.6 mM.  $\bullet$ , HbO<sub>2</sub> + CN<sup>-</sup>, pH 7.4; heme concentration, 11.1 mM; CN<sup>-</sup>, 2 mM; equilibrated with 1 atm O<sub>2</sub>. —, best-fit curve for a Curie law paramagnetic component plus the contraction from a coupled spin pair with S<sub>1</sub> = S<sub>2</sub> =  $\frac{1}{2}$ , g = 2.00, TIP = 0, and J = -72 cm<sup>-1</sup> for HbO<sub>2</sub> or J = -73.5 cm<sup>-1</sup> for HbO<sub>2</sub> + CN<sup>-</sup>.  $\bullet$  HbCO equilibrated with 1 atm CO, pH 7.2; heme concentration, 10.6 mM.

When we attempted a similar fit for the HbCO data, we only found a lower limit in the form  $|J_{CO}| \ge 400 \text{ cm}^{-1}$ .

Data on native  $HbO_2$  and on stripped  $HbO_2$  containing  $CN^-$  gave similar results, with  $J=-72.5~cm^{-1}$  and  $J=-73.5~cm^{-1}$ , respectively. The differences in J among the three samples are not significant in the resolution of our fitting procedure. The paramagnetic component was larger in the sample containing  $CN^-$  because the sample had been equilibrated with pure  $O_2$ 

rather than with air. When this is accounted for, the residual MetHb concentration is similar for both oxygenated samples, about 3%.

The measurements on stripped HbO<sub>2</sub> with added CN<sup>-</sup> were performed because CN<sup>-</sup> is known to bind strongly to MetHb, stabilizing it in a low-spin state within the whole temperature range explored (15). Thus, no composite temperature-dependent behavior can be expected from any MetHb present in the

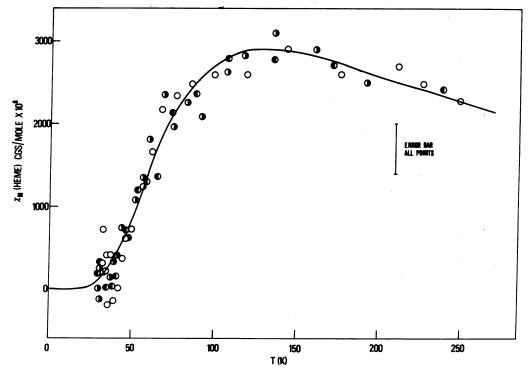


FIG. 2. Deviation from Curie law and diamagnetic background for the oxygenated samples, expressed as the dependence of the suceptibility per heme as extracted from the display of data of Fig. 1, and for native HbO<sub>2</sub>. O, stripped HbO<sub>2</sub>. ( $\bullet$ ), stripped HbO<sub>2</sub> + CN<sup>-</sup>.  $\bullet$ , native HbO<sub>2</sub>; 11.2 mM in heme, pH 7.2, equilibrated with 1 atm O<sub>2</sub>. —, theoretical curve for S<sub>1</sub> = S<sub>2</sub> = ½, g = 2.00, TIP = 0, and J = -73 cm<sup>-1</sup>.

sample. The agreement between the data for the CN<sup>-</sup> sample and the other data is taken as a crucial test that the temperature-dependent behavior observed in HbO<sub>2</sub> solutions is indeed due to the iron-oxygen complex. We can also rule out a temperature-dependent dissociation of diamagnetic HbO<sub>2</sub> to deoxyHb plus O<sub>2</sub>. Indeed, we obtained the same results for the nonlinear component, while the oxygen affinity increased by at least a factor of 5 on going from native to stripped Hb samples and changing the oxygen concentration by a factor of 5 (equilibrating with air or with pure O<sub>2</sub>) (see also Fig. 2). Moreover, no sign of dissociation could be detected in the optical spectra.

To emphasize the agreement between the three oxygenated samples and the satisfactory fit with the theoretical curves, we show in Fig. 2 the experimental data for all three kinds of samples expressed as molar susceptibility per heme, after subtraction of the diamagnetism of the solution and of the Curie law paramagnetic component, as given by the computer fitting procedure. It is apparent that the characteristic nonlinear behavior of the atomic susceptibility is due to a triplet state that becomes thermally populated from a singlet state. The full curve represents the corresponding theoretical curve with  $J = -73 \text{ cm}^{-1}$ , which fits well (within experimental error) all the data from different samples.

From our experimental evidence we conclude that, at variance with what is widely accepted,  $HbO_2$  is not diamagnetic above about 50 K and that the iron-oxygen complex contains a coupled  $S = \frac{1}{2}$  spin pair.

In regard to the electronic state of the iron-oxygen bond, as suggested by the present data, we notice that an  $S = \frac{1}{2}$  pair in the bond would be consistent with the assignment by Barlow et al. (18) of a bond order of 1.5 for the O-O bond in HbO<sub>2</sub>. It is also in accord with the enhanced O<sub>2</sub> uptake by model iron porphyrins in polar aprotic solvents (19, 20). If we assign one electron to the O-O bond and the other to the Fe-O bond, our data require an antiferromagnetic coupling between their spins. This is consistent with the Weiss model (4), which gives  $Fe^{3+}O_2^{-}$ . Whatever the localization of these two electrons, the iron would still be low-spin, with at most one unpaired electron. This would still agree with the view of the iron being in the low-spin state when ligated (21).

We are grateful to H. B. Gray for encouragement and discussions during this work and we also thank D. Dooley for discussions. We are indebted to R. A. Noble for suggesting the test with CN<sup>-</sup>, to Max Perutz for encouragement and for a critical reading of the manuscript, and to M. Berardo and E. Gori for continuous technical assistance with the cryogenics of the magnetometer. This work was supported by Gruppo Nazionale Struttura della Materia of the Consiglio Nazionale delle Ricerche.

- Pauling, L. & Coryell, C. D. (1936) Proc. Natl. Acad. Sci. USA 22, 210-216.
- Pauling, L. (1949) Hemoglobin (Butterworth Sci. Publ., London), pp. 57–65.
- 3. Griffith, J. S. (1956) Proc. R. Soc. London Ser. A 235, 23-36.
- Weiss, J. J. (1964) Nature 202, 83–84.
- 5. Gray, H. B. (1971) Adv. Chem. Ser. 100, 365-389.
- Goddard, W., & Olafson, B. D. (1975) Proc. Natl. Acad. Sci. USA 72, 2335–2339.
- 7. Koster, A. S. (1972) J. Chem. Phys. 56, 3161-3162.
- Wittenberg, J. D., Wittenberg, B. A., Peisach, J. & Blumberg, W. E. (1970) Proc. Natl. Acad. Sci. USA 67, 1846-1852.
- Hoffman, B. M., Diamante, D. L. & Basolo, F. (1970) J. Am. Chem. Soc. 92, 61-65.
- 10. Lang, G. & Marshall, W. (1966) J. Mol. Biol. 18, 358-404.
- Spiro, T. G. & Strekas, T. C. (1974) J. Am. Chem. Soc. 96, 338-345.
- Cerdonio, M., Mogno, F., Romani, G. L., Messana, C. & Gramacciomi, C. (1977) Rev. Sci. Instrum. 48, 19-25.
- Cerdonio, M., Cosmelli, C., Romani, G. L., Messana, C. & Gramaccioni, C. (1976) Rev. Sci. Instrum. 47, 1-5.
- Benesch, R., Benesch, R. E. & Yu, C. K. (1968) Proc. Natl. Acad. Sci. USA 59, 526–530.
- Iizuka, T. & Kotani, M. (1969) Biochim. Biophys. Acta 194, 351-363.
- Havemann, R., Habeditzl, W. & Rabe, G. (1962) Z. Phys. Chem. 218, 419–425.
- Mabbs, F. E. & Machin, D. J. (1973) Magnetism and Transition Metal Complexes (Chapman and Hall ed., London).
- Barlow, C. H., Maxwell, J. C., Wallace, W. J. & Caughey, W. S. (1973) Biochem. Biophys. Res. Commun. 55, 91-95.
- Anderson, D. L., Weschler, C. J. & Basolo, F. (1974) J. Am. Chem. Soc. 96, 5599-5600.
- Basolo, F., Hoffman, B. M. & Ibers, J. A. (1975) Acc. Chem. Res. 8, 384-392.
- 21. Perutz, M. F. (1970) Nature 228, 726-739.